

Single-Polynucleotide Lipid Nanoparticles: Ultrafast Assembly and Multiscale Structure

Maxime Guichonnet¹, Johanna Mirande-Ney¹, Sébastien Teychene¹ and Kevin Roger¹

¹ Laboratoire de Génie Chimique, Université de Toulouse, CNRS, Institut National Polytechnique de Toulouse, Toulouse 31400, France

maxime.guichonnet@toulouse-inp.fr

Lipid nanoparticles (LNPs) have emerged as key carriers for the delivery of polynucleotides such as mRNA, and their two-step synthesis involving nanoprecipitation followed by dialysis is under intense scrutiny in the scientific literature. Despite their widespread use, the mechanisms governing LNP formation remain poorly understood, involving complex internal structures, multiple particle populations, and strong coupling between composition, assembly pathway, and delivery performance. This complexity makes it difficult to establish clear structure–function relationships and calls for approaches yielding more homogeneous and well-defined objects. Here, we show that LNP assembly can be directed toward the formation of smaller and more structurally defined nanoparticles. Using small-angle X-ray and neutron scattering (SAXS/SANS) and cryogenic transmission electron microscopy (cryo-TEM), we demonstrate that the resulting nanoparticles encapsulate a single polynucleotide and quantify their multiscale organization. Direct in situ measurements reveal that these particles assemble through a rapid and cooperative co-assembly process occurring on the millisecond timescale^{1,2}, consistent with a kinetically driven pathway far from equilibrium controlled by molecular diffusion rather than colloidal aggregation³. We further show that the final structure is highly responsive to its environment, including ionic strength and pH, highlighting the role of physicochemical conditions in tuning nanoparticle organization^{3–5}. Despite this responsiveness, the nanoparticles remain stable upon storage at 4 °C while maintaining efficient transfection. Overall, this work establishes a direct link between assembly pathway, structure, and function, and provides new insights for the rational design of lipid-based nanocarriers.

Keywords:

Lipid nanoparticle, kinetics, co-assembly

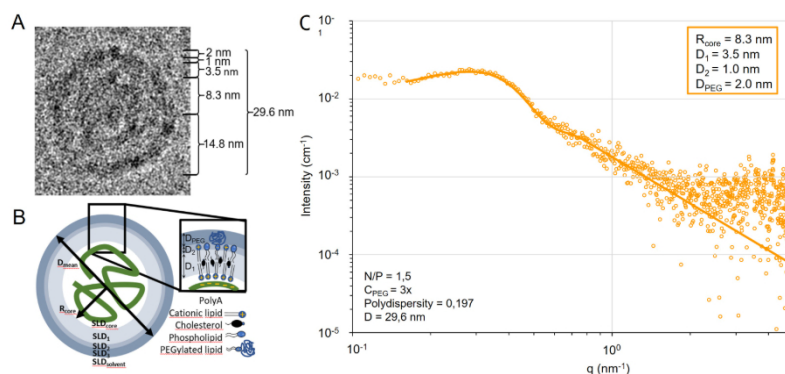


Fig. 1: (A) Representative cryo-TEM image of PEGylated LNP, revealing the internal structural features and characteristic length scales within the particles. (B) Proposed structural model of the LNP architecture. (C) SAXS profile of PEGylated LNPs fitted using the structural model shown in panel (B)

References:

- (1) Schober, G. B.; Story, S.; Arya, D. P. A Careful Look at Lipid Nanoparticle Characterization: Analysis of Benchmark Formulations for Encapsulation of RNA Cargo Size Gradient. *Sci Rep* **2024**, *14* (1), 2403.
- (2) Zhigaltsev, I. V.; Belliveau, N.; Hafez, I.; Leung, A. K. K.; Huft, J.; Hansen, C.; Cullis, P. R. Bottom-Up Design and Synthesis of Limit Size Lipid Nanoparticle Systems with Aqueous and Triglyceride Cores Using Millisecond Microfluidic Mixing. *Langmuir* **2012**, *28* (7), 3633–3640.
- (3) Chen, D.; Liu, Z.; Guo, L.; Yang, L.; Zhao, Y.; Yang, M. Controlled Preparation of Lipid Nanoparticles in Microreactors: Mixing Time, Morphology and mRNA Delivery. *Chemical Engineering Journal* **2025**, *505*, 159318.
- (4) Binici, B.; Rattray, Z.; Zinger, A.; Perrie, Y. Exploring the Impact of Commonly Used Ionizable and Pegylated Lipids on mRNA-LNPs: A Combined *in Vitro* and Preclinical Perspective. *Journal of Controlled Release* **2025**, *377*, 162–173.
- (5) Li, Z.; Carter, J.; Santos, L.; Webster, C.; van der Walle, C. F.; Li, P.; Rogers, S. E.; Lu, J. R. Acidification-Induced Structure Evolution of Lipid Nanoparticles Correlates with Their In Vitro Gene Transfections. *ACS Nano* **2023**, *17* (2), 979–990.